

# TransIT-HeLaMONSTER<sup>®</sup> Transfection Kit

Protocol for MIR 2900, 2904, 2905, 2906



The Transfection Experts

Quick Reference Protocol, MSDS and Certificate of Analysis available at [mirusbio.com/2900](http://mirusbio.com/2900)

## INTRODUCTION

TransIT-HeLaMONSTER<sup>®</sup> Transfection Kit is specifically optimized to provide exceptional transfection efficiency of plasmid DNA in HeLa cells and cell types of associated lineage. TransIT-HeLaMONSTER Transfection Kit contains two components, namely: TransIT<sup>®</sup>-HeLa Reagent and the MONSTER Reagent. This kit provides all the attributes of the trusted TransIT series of transfection reagents: high transfection efficiency, low toxicity, serum compatibility, simplicity of use and reproducibility. Transfection with this kit does not require medium changes and can be carried out in serum-containing medium. This kit is suitable for both transient and stable transfection.

## SPECIFICATIONS

|                          |  |
|--------------------------|--|
| <b>Storage</b>           | Store both TransIT-HeLa Reagent and MONSTER Reagent at -20°C. <b>Before each use</b> , warm to room temperature and vortex gently. |
| <b>Product Guarantee</b> | 1 year from the date of purchase, when properly stored and handled.  |



Warm TransIT-HeLa and MONSTER Reagent to room temperature and vortex gently before each use.

## MATERIALS

### Materials Supplied

The TransIT-HeLaMONSTER Transfection Kit is supplied in *one* of the following formats.

| Product No. | Volume of TransIT-HeLa Reagent | Volume of MONSTER Reagent |
|-------------|--------------------------------|---------------------------|
| MIR 2904    | 1 × 0.4 ml                     | 1 × 0.4 ml                |
| MIR 2900    | 1 × 1 ml                       | 1 × 1 ml                  |
| MIR 2905    | 5 × 1 ml                       | 5 × 1 ml                  |
| MIR 2906    | 10 × 1 ml                      | 10 × 1 ml                 |

### Materials required, but not supplied

- Cultured cells
- Appropriate cell culture medium
- Purified DNA
- Serum-free medium (e.g. Opti-MEM<sup>®</sup> I Reduced-Serum Medium)
- Sterile tube for transfection complex preparation
- Micropipets
- Reporter assay as required

**For Research Use Only.**

## BEFORE YOU START:

### Important Tips for Optimal DNA Transfection

Optimize reaction conditions for each HeLa cell subtype to ensure successful transfections. The suggestions below yield high efficiency DNA transfection using the *TransIT-HeLaMONSTER* Transfection Kit. **Tables 1–2** on Pages 3–4 present recommended starting conditions depending on culture vessel size.

- **Cell density (% confluence) at transfection.** The recommended cell density for HeLa cell subtypes at transfection is  $\geq 80\%$  confluence. Determine the optimal cell density for each HeLa cell subtype in order to maximize transfection efficiency. Divide the cells 18–24 hours before transfection to ensure that the cells are actively dividing and reach the appropriate cell density at the time of transfection.
- **DNA purity.** Use highly purified, sterile, and contaminant-free DNA for transfection. Plasmid DNA preps that are endotoxin-free and have  $A_{260/280}$  absorbance ratio of 1.8–2.0 are desirable. DNA prepared using miniprep kits is not recommended as it may contain high levels of endotoxin. We recommend using MiraCLEAN® Endotoxin Removal Kit (MIR 5900) to remove any traces of endotoxin from your DNA preparation.
- **TransIT-HeLa Reagent:DNA ratio.** As a starting point, use 3  $\mu\text{l}$  of *TransIT-HeLa* Reagent per 1  $\mu\text{g}$  of DNA. The optimal *TransIT-HeLa* Reagent to DNA ratio can be determined by titrating the reagent from 2–4  $\mu\text{l}$  per 1  $\mu\text{g}$  of DNA. Please refer to Tables 1–2 on Pages 3–4 for recommended starting conditions.
- **MONSTER Reagent:DNA ratio.** Different MONSTER Reagent amounts may be required depending on the cell culture and experimental conditions. The optimal MONSTER Reagent:DNA ratio should be determined by titrating the reagent from 0–5  $\mu\text{l}$  per 1  $\mu\text{g}$  of DNA. Please refer to **Tables 1–2** on Pages 3–4 for recommended starting conditions.
- **Complex formation conditions.** Prepare *TransIT-HeLa*:MONSTER:DNA complexes in serum-free growth medium. Mirus recommends Opti-MEM I Reduced-Serum Medium.
- **Cell culture conditions:** Culture cells in the appropriate medium, with or without serum. There is no need to perform a medium change to remove the transfection complexes. *TransIT-HeLaMONSTER* Transfection Kit yields improved efficiencies when transfections are performed in complete growth medium (instead of serum-free medium) without a post-transfection medium change.
- **Presence of antibiotics:** Antibiotics will inhibit transfection complex formation and therefore should be excluded from the complex formation step. Transfection complexes can be added to cells grown in complete culture medium containing low levels of antibiotics (0.1–1X final concentration of penicillin/streptomycin mixture).
- **Post-transfection incubation time.** Determine the best incubation time post-transfection for each cell type. The optimal incubation time is generally 24–72 hours, but will vary depending on the goal of the experiment, nature of the plasmid, and the half-life of the expressed protein.



**Do not** use DNA prepared using miniprep kits for transfection.



**Do not** use serum or antibiotics in the medium during transfection complex formation.

## DNA TRANSFECTION PROTOCOL

The following procedure describes how to perform DNA transfections in 6-well plates. The surface areas of other culture vessels are different and transfections must be scaled accordingly. Appropriately increase or decrease the amounts of serum free medium, *TransIT-HeLa* Reagent, MONSTER Reagent, DNA and complete culture medium based on the surface area of the cell culture vessel. **Table 1** presents recommended starting conditions depending on culture vessel size.

Optimal transfection efficiencies can be achieved using the following protocol. An alternate protocol (please refer to Page 4) is provided which uses a slightly modified procedure to apply diluted MONSTER Reagent directly to the cells. Choose the protocol that achieves the highest transfection efficiency.

**Table 1.** Recommended starting conditions for DNA transfections with the *TransIT-HeLaMONSTER* Transfection Kit.

| Culture vessel              | 96-well plate        | 48-well plate       | 24-well plate       | 12-well plate       | 6-well plate        | 10-cm dish         | T75 flask          |
|-----------------------------|----------------------|---------------------|---------------------|---------------------|---------------------|--------------------|--------------------|
| Surface area                | 0.35 cm <sup>2</sup> | 1.0 cm <sup>2</sup> | 1.9 cm <sup>2</sup> | 3.8 cm <sup>2</sup> | 9.6 cm <sup>2</sup> | 59 cm <sup>2</sup> | 75 cm <sup>2</sup> |
| Complete growth medium      | 92 µl                | 263 µl              | 0.5 ml              | 1.0 ml              | 2.5 ml              | 15.5 ml            | 19.7 ml            |
| Serum-free medium           | 9 µl                 | 26 µl               | 50 µl               | 100 µl              | 250 µl              | 1.5 ml             | 1.9 ml             |
| DNA (1µg/µl stock)          | 0.1 µl               | 0.25 µl             | 0.5 µl              | 1 µl                | 2.5 µl              | 15.5 µl            | 19.7 µl            |
| <i>TransIT-HeLa</i> Reagent | 0.3 µl               | 0.75 µl             | 1.5 µl              | 3 µl                | 7.5 µl              | 46.5 µl            | 59.1 µl            |
| MONSTER Reagent*            | 0.2 µl               | 0.50 µl             | 1 µl                | 2 µl                | 5 µl                | 31 µl              | 39.4 µl            |

\*Different MONSTER Reagent amounts may be required depending on the cell culture and experimental conditions. The optimal MONSTER Reagent:DNA ratio should be determined by titrating the reagent from 0–5 µl per 1 µg of DNA.



Surface areas are based on Greiner tissue culture plates and Falcon 10-cm dishes and T75 flasks. All volumes given are per well (or per dish) for a given culture vessel.

If small volumes of kit reagents need to be pipetted, dilute the *TransIT-HeLa* and MONSTER reagent in serum-free medium and water, respectively. **Do not** store diluted reagents.

### Transient DNA transfection protocol per well of a 6-well plate

#### A. Plate cells

1. Approximately 18–24 hours before transfection, plate cells in 2.5 ml complete growth medium per well in a 6-well plate. Ideally cells should be ≥80% confluent prior to transfection.
2. Incubate the cell cultures overnight.

#### B. Prepare *TransIT-HeLa*:MONSTER:DNA complexes (Immediately before transfection)

1. Warm *TransIT-HeLa* and MONSTER reagents to room temperature and vortex gently before using.
2. Place 250 µl of Opti-MEM I Reduced-Serum Medium in a sterile tube.
3. Add 2.5 µg (2.5 µl of a 1 µg/µl stock) DNA. Pipet gently to mix completely.
4. Add 7.5 µl *TransIT-HeLa* Reagent to the diluted DNA mixture. Pipet gently to mix completely.
5. Add 5 µl MONSTER Reagent to the diluted *TransIT-HeLa*:DNA mixture. Pipet gently to mix completely.



Divide cultured cells 18–24 hours before transfection to ensure active cell division at the time of transfection.



Warm *TransIT-HeLa* and MONSTER reagents to room temperature and vortex gently before each use.

6. Incubate at room temperature for 15–30 minutes to allow sufficient time for complexes to form.
7. Add 2.5 ml (per well) of pre-warmed complete growth medium to the complexes in the tube.

**C. Distribute the complexes to cells in complete growth medium**

1. Remove the medium from the cells immediately prior to the addition of the complexes.
2. Add the *TransIT-HeLa:MONSTER:DNA* complexes drop-wise to different areas of the wells. It is not necessary to replace the complete growth medium with fresh medium.
3. Gently rock the culture vessel back-and-forth and from side-to-side to evenly distribute the *TransIT-HeLa:MONSTER:DNA* complexes.
4. Incubate for 24–72 hours. It is not necessary to replace the complete growth medium with fresh medium.
5. Harvest cells and assay as required.



There is no need to change culture medium after transfection. If required, perform a medium change at least 4 hours post-transfection.

For generating stable cell transfectants, passage the cells 48-72 hours post-transfection in complete growth medium containing the appropriate selection antibiotic such as G418 or Hygromycin B. Maintain selection for 1–2 weeks, allowing selection of cells that have undergone stable integration of DNA.

**Alternate transient DNA transfection protocol per well of a 6-well plate**

The following *TransIT-HeLaMONSTER* Transfection Kit protocol uses a slightly modified procedure where diluted *MONSTER* Reagent is applied directly to the cells plated in a 6-well plate. In some cases, this protocol might yield higher transfection efficiency compared to the standard protocol. The surface areas of other culture vessels are different and transfections must be scaled accordingly. Appropriately increase or decrease the amounts of serum free medium, *TransIT-HeLa* Reagent, *MONSTER* Reagent, DNA and complete culture medium based on the surface area of the cell culture vessel. **Table 2** presents recommended starting conditions depending on culture vessel size.

**Table 2.** Recommended starting conditions for DNA transfections with the *TransIT-HeLaMONSTER* Transfection Kit using the alternate protocol.

| Culture vessel                        | 96-well plate        | 48-well plate       | 24-well plate       | 12-well plate       | 6-well plate        | 10-cm dish         | T75 flask          |
|---------------------------------------|----------------------|---------------------|---------------------|---------------------|---------------------|--------------------|--------------------|
| Surface area                          | 0.35 cm <sup>2</sup> | 1.0 cm <sup>2</sup> | 1.9 cm <sup>2</sup> | 3.8 cm <sup>2</sup> | 9.6 cm <sup>2</sup> | 59 cm <sup>2</sup> | 75 cm <sup>2</sup> |
| Complete growth medium                | 92 µl                | 263 µl              | 0.5 ml              | 1.0 ml              | 2.5 ml              | 15.5 ml            | 19.7 ml            |
| Serum-free medium                     | 9 µl                 | 26 µl               | 50 µl               | 100 µl              | 250 µl              | 1.5 ml             | 1.9 ml             |
| DNA (1µg/µl stock)                    | 0.1 µl               | 0.25 µl             | 0.5 µl              | 1 µl                | 2.5 µl              | 15.5 µl            | 19.7 µl            |
| <i>TransIT-HeLa</i> Reagent           | 0.3 µl               | 0.75 µl             | 1.5 µl              | 3 µl                | 7.5 µl              | 46.5 µl            | 59.1 µl            |
| Diluted <i>MONSTER</i> Reagent (0.1X) | 2.0 µl               | 5.0 µl              | 10 µl               | 20 µl               | 50 µl               | 310 µl             | 394 µl             |

**A. Plate cells**

1. Approximately 18–24 hours before transfection, plate cells in 2.5 ml complete growth medium per well in a 6-well plate. Ideally cells should be ≥80% confluent (~1–3 × 10<sup>5</sup> cells/well) prior to transfection.
2. Incubate the cell cultures overnight.



If small volumes of kit reagents need to be pipetted, dilute the *TransIT-HeLa* and *MONSTER* reagent in serum-free medium and water, respectively. **Do not** store diluted reagents.



Divide cultured cells 18–24 hours before transfection to ensure active cell division at the time of transfection.

**B. Prepare *TransIT-HeLa*:DNA complexes****(Immediately before transfection)**

1. Warm *TransIT-HeLa* Reagent to room temperature and vortex gently before using.
2. Place 250  $\mu$ l of Opti-MEM I Reduced-Serum Medium in a sterile tube.
3. Add 2.5  $\mu$ g (2.5  $\mu$ l of a 1  $\mu$ g/ $\mu$ l stock) DNA. Pipet gently to mix completely.
4. Add 7.5  $\mu$ l *TransIT-HeLa* Reagent to the diluted DNA mixture. Pipet gently to mix completely.
5. Incubate at room temperature for 15–30 minutes to allow sufficient time for complexes to form.

**C. Dilution of MONSTER Reagent****(Immediately prior to or during *TransIT-HeLa*:DNA complex formation)**

1. Warm MONSTER Reagent to room temperature and vortex gently before using.
2. In a sterile tube, dilute 5  $\mu$ l MONSTER Reagent 10-fold in 45  $\mu$ l sterile (DNase and RNase-free) water. Mix well.
3. Immediately return the stock MONSTER Reagent to -20°C after use.

**D. Distribute the complexes to cells in complete growth medium**

1. If necessary, remove the medium from the cells prepared in Step A and replace with 2.5 ml per well of fresh complete growth medium.
2. Add the *TransIT-HeLa*:DNA complexes drop-wise to different areas of the wells.
3. Gently rock the culture vessel back-and-forth and from side-to-side to evenly distribute the *TransIT-HeLa*:DNA complexes.
4. Add the diluted MONSTER Reagent prepared in Step C drop-wise directly to the cells.
5. Gently rock the culture vessel back-and-forth and from side-to-side to evenly distribute the diluted MONSTER Reagent.
6. Incubate for 24–72 hours. It is not necessary to replace the complete growth medium with fresh medium.
7. Harvest cells and assay as required.



Warm *TransIT-HeLa* reagent to room temperature and vortex gently before each use.



There is no need to change culture medium after transfection. If required, perform a medium change at least 4 hours post-transfection.

For generating stable cell transfectants, passage the cells 48–72 hours post-transfection in complete growth medium containing the appropriate selection antibiotic such as G418 or Hygromycin B. Maintain selection for 1–2 weeks, allowing selection of cells that have undergone stable integration of DNA.

## TROUBLESHOOTING GUIDE

| Problem  | Solution   |
|--|--|
| <b>LOW DNA TRANSFECTION EFFICIENCY</b>                         |  |
| <i>TransIT-HeLa</i> or MONSTER Reagent was not mixed properly. | Warm <i>TransIT-HeLa</i> and MONSTER Reagents to room temperature and vortex gently before each use.   |
| Suboptimal amount of <i>TransIT-HeLa</i> Reagent               | Determine optimal amount of <i>TransIT-HeLa</i> Reagent for each HeLa cell subtype. Titrate the <i>TransIT-HeLa</i> Reagent from 2–4 µl per 1 µg DNA. Refer to “Before You Start” on Page 2.   |
| Suboptimal amount of MONSTER Reagent                           | Determine optimal amount of MONSTER Reagent for each HeLa cell subtype. Titrate the MONSTER Reagent from 0–5 µl per 1 µg DNA. Refer to “Before You Start” on Page 2.   |
| Suboptimal DNA concentration                                   | <p>Confirm DNA concentration and purity. Use plasmid DNA preps that have an <math>A_{260/280}</math> absorbance ratio of 1.8–2.0.</p> <p>The optimal DNA concentration generally ranges between 1–3 µg/well of a 6-well plate. Start with 2.5 µg/well of a 6-well plate. Consider testing more or less DNA while scaling the amount of <i>TransIT-HeLa</i> and MONSTER Reagents accordingly.</p>   |
| Low-quality plasmid DNA  | <p>Use highly purified, sterile, endotoxin and contaminant-free DNA for transfection.</p> <p>We recommend using Mirus Bio’s MiraCLEAN Endotoxin Removal Kit (MIR 5900) for removal of endotoxin from your DNA preparation.</p> <p>Alternatively, use cesium chloride gradient or anion exchange purified DNA which contains levels of endotoxin that do not harm most cells.</p> <p><b>Do not</b> use DNA prepared using miniprep kits as it may contain high levels of endotoxin.</p>   |
| Inhibitor present during transfection                          | <p>Serum and antibiotics inhibit transfection complex formation. Prepare transfection complexes in serum-free growth medium. We recommend Opti-MEMI Reduced-Serum Medium. Once transfection complexes are formed, they can be added directly to cells cultured in complete growth medium containing serum and 0.1–1X antibiotics.</p> <p>Polyanions such as dextran sulfate or heparin can inhibit transfection. Use culture medium that does not contain these polyanions. If necessary, the transfection medium can be replaced with polyanion containing medium 24 hours post transfection.</p> |
| Incorrect vector sequence                                      | If you do not observe expression of your target insert, verify the sequence of the plasmid DNA.  |
| Transfection incubation time                                   | Determine the optimal transfection incubation time for each cell type and experiment. Test a range of incubation times (e.g. 12–72 hours). The best incubation time is generally 24–48 hours.  |
| Cells not actively dividing at the time of transfection        | Divide the culture at least 18–24 hours before transfection to ensure that the cells are actively dividing and reach optimal cell density at time of transfection.   |
| Precipitate formation during transfection complex formation    | <p>During complex formation, scale all reagents including serum-free medium, <i>TransIT-HeLa</i> Reagent, MONSTER Reagent and plasmid DNA according to Table 1–2 on Pages 3–4.</p> <p>Precipitation may be observed when excess DNA is used during complex formation. This may negatively impact transfection efficiency. To avoid precipitation when using high concentrations of DNA, increase the volume of serum-free medium during complex formation by two-fold.</p>   |

## TROUBLESHOOTING GUIDE continued

| Problem  | Solution   |
|--|--|
| <b>LOW DNA TRANSFECTION EFFICIENCY</b>                                       |  |
| Proper experimental controls were not included                               | <p>To verify efficient transfection, use <i>TransIT-HeLaMONSTER</i> Transfection Kit to deliver a positive control such as a luciferase, beta-galactosidase or green fluorescent protein (GFP) encoding plasmid.</p> <p>To assess delivery efficiency of plasmid DNA, use Mirus' <i>Label IT® Tracker™</i> Intracellular Nucleic Acid Localization Kit to label the target plasmid or Mirus' pre-labeled <i>Label IT Plasmid Delivery Controls</i> (please refer to Related Products on Page 8).</p> |
| <b>HIGH CELLULAR TOXICITY</b>  |  |
| Transfection complexes and cells not mixed thoroughly after complex addition | Add transfection complexes drop-wise to different areas of the wells containing plated cells . Gently rock the dish back-and-forth and from side-to-side to distribute the complexes evenly. Do not swirl or rotate the dish, as this may cause uneven distribution.   |
| Transfection complexes added to cells cultured in serum-free medium          | Allow transfection complexes to form in serum-free medium, then add these complexes to cells cultured in complete growth medium. The presence of serum in the growth medium improves transfection efficiency and reduces cytotoxicity. No culture medium change is required after the addition of transfection complexes to cells.   |
| Endotoxin-contaminated plasmid DNA   | Use highly purified, sterile, endotoxin and contaminant-free DNA for transfection.   |
|  | We recommend using Mirus Bio's <i>MiraCLEAN</i> Endotoxin Removal Kit (MIR 5900) for removal of any traces of endotoxin from your DNA preparation. Alternatively, use cesium chloride gradient or anion exchange purified DNA which contains levels of endotoxin that do not harm most cells.  |
| Expressed target gene is toxic to cells                                      | Do not use DNA prepared using miniprep kits as it may contain high levels of endotoxin.  |
|  | Compare toxicity levels against a cells alone control and cells transfected with an empty vector to assess the cytotoxic effects of the target protein being expressed.  |
| Cell density not optimal at time of transfection                             | If lower levels of target gene expression are desired in your transfection experiments, consider reducing the amount of target plasmid. Maintain the optimal <i>TransIT-HeLaMONSTER</i> :DNA ratio by using carrier DNA such as an empty cloning vector.   |
|  | Determine the best cell density for each HeLa cell subtype to maximize transfection efficiency. Use this cell density in subsequent experiments to ensure reproducibility. For most HeLa cell subtypes, ≥80% confluence is recommended at transfection, but use of higher or lower densities may increase cell viability depending on cell type.   |
| Cell morphology has changed  | Mycoplasma contamination can alter cell morphology and affect transfection efficiency. Check your cells for Mycoplasma contamination. Use a fresh frozen stock of cells or use appropriate antibiotics to eliminate Mycoplasma.  |
|  | A high or low cell passage number can make cells more sensitive and refractory to transfection. Maintain a similar passage number between experiments to ensure reproducibility.   |

## RELATED PRODUCTS

- Ingenio<sup>®</sup> Electroporation Solution and Kits
- Label IT<sup>®</sup> Plasmid Delivery Controls
- Label IT<sup>®</sup> Tracker™ Intracellular Nucleic Acid Localization Kits
- MiraCLEAN<sup>®</sup> Endotoxin Removal Kits
- TransIT-X2<sup>®</sup> Dynamic Delivery System
- TransIT<sup>®</sup>-3D Transfection Reagent
- TransIT<sup>®</sup>-2020 Transfection Reagent
- TransIT-PRO<sup>®</sup> Transfection Kit
- TransIT<sup>®</sup>-LT1 Transfection Reagent
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Contact Mirus Bio for additional information.



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